THE STRUCTURE OF A NEW ISOTETRACENONE ANTIBIOTIC, CAPOAMYCIN

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Summary: Based on  $^{1}$ H and  $^{13}$ C NMR spectral data and chemical degradation, the structure of capoamycin, a new isotetracenone antibiotic has been determined as shown in Fig. 1.

Capoamycin is a new antitumor antibiotic produced by <u>Streptomyces capoamus</u> No. 23-41, which belongs to the isotetracenone antibiotic group.<sup>1)</sup> This substance induced differentiation of mouse myeloid leukemia cells  $(M1)^{2}$  and prolonged the survival periods of mice bearing Ehrlich ascites carcinoma.

The physicochemical properties of capoamycin (<u>1</u>) are as follows: orange yellow powder, no definite m.p.; FAB-MS,  $\underline{m}/\underline{z}$  724 (M + diethanolamine + H)<sup>+</sup>; <u>Anal</u>. found, C 68.03, H 6.26, O 25.71, calcd. for  $C_{35}H_{38}O_{10}$ , C 67.95, H 6.19, O 25.86;  $[\alpha]_D^{21}$  +209°(c 0.1, acetone); UV  $\lambda \underset{max}{\text{MeOH}}$  nm ( $E_{1\text{cm}}^{1\text{K}}$ ), 235(704), 257(782), 305(139) and 442(100). The IR spectrum of <u>1</u> (KBr) showed the absorptions due to hydroxyl groups (3420 cm<sup>-1</sup>), an unsaturated ester (1710 cm<sup>-1</sup>), an unsaturated ketone in a 6-membered ring (1690 cm<sup>-1</sup>), a non-chelated quinone (1670 cm<sup>-1</sup>, sh) and a chelated quinone carbonyl group (1638 cm<sup>-1</sup>).



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The 400 MHz <sup>1</sup>H NMR spectrum<sup>3</sup>) of <u>1</u> in CDCl<sub>3</sub> revealed the following signals:  $\delta$  12.25 (s, exchangeable, 8-OH), 7.84 (d, J=7.6 Hz, 10-H), 7.58 (d, J=7.6, 11-H), 6.91 (d, J=10.0, 6-H), 6.43 (d, J=10.0, 5-H), 6.22 (br.s, 2-H), 2.76 (d, J=18.8, 4-Ha), 2.51 (d, J=18.8, 4-Hb) and 1.94 (3H, s, 3-CH<sub>3</sub>) ascribed to a modified benz[a]anthraquinone chromophore which is characteristic to the isotetracenone antibiotics;  $\delta$  4.87 (dd, J=11.2, 1.6, 1'-H), 4.64 (dd, J=9.4, 9.0, 4'-H), 3.97 (ddd, J=11.0, 9.0, 5.3, 3'-H), 3.66 (dq, J=9.4, 6.0, 5'-H), 2.57 (ddd, J=11.3, 5.3, 1.6, 2'-Heq), 1.51 (ddd, J=11.3, 11.2, 11.1, 2'-Hax) and 1.30 (3H, d, J=6.0, 6'-H) assignable to a  $\beta$ -olivoside moiety;  $\delta$  7.33 (m, 3"-H), 6.19 (2H, m, 4"-H and 5"-H), 5.87 (d, J=15.4, 2"-H), 2.19 (2H, dt, J=5.4, 7.1, 6"-H<sub>2</sub>), 1.44 (2H, tt, J=7.3, 7.1, 7"-H<sub>2</sub>), 1.31 (4H, m, 8"-H<sub>2</sub> and 9"-H<sub>2</sub>) and 0.90 (3H, t, J=7.4, 10"-H<sub>3</sub>) due to a 2.4-decadienoic acid side chain.

Based on  $\{{}^{13}C- {}^{1}H\}$  selective proton decoupling experiments<sup>3)</sup> of <u>1</u> at 100 MHz, the following signals have been assigned (see Fig. 1):  $\delta$  78.8 (d, C-4'), 74.3 (d, C-5'), 71.5 (d, C-3'), 71.2 (d, C-1'), 40.1 (t, C-2') and 18.4 (q, C-6') due to the olivose residue; 167.7 (s, C-1"), 146.5 (d, C-3"), 145.8 (d, C-5"), 127.9 (d, C-4"), 117.9 (d, C-2"), 33.2 (t, C-6"), 31.6 (t, C-8"), 28.6 (t, C-7"), 22.7 (t, C-9") and 14.3 (q, C-10") due to the 2,4-decadienoic acid moiety. Long range selective proton decoupling (LSPD) experiments established the tetracyclic structure and the  ${}^{13}C$  chemical shift assignments of the chromophore moiety as shown in Fig.2. The linkage of C and D rings was proved by observing nuclear Overhauser effect between 4-H<sub>2</sub> and 5-H. Thus, the structure of the chromophore was established to be 4a,12b-dihydro-4a,8,12b-trihydroxy-3-methyl-benz[a]anthracene-1,7,12(4H)-trione.



The long range coupling between 10-H and 1'-H indicates that the olivose residue is connected to C-9 position with a C-glycosidic linkage.

Hydrolysis of <u>1</u> with 0.1 N NaOH at room temperature for 30 min gave a rearranged chromophore with a C-olivoside (<u>2</u>)  $[C_{25}H_{22}O_8; FAB-MS, \underline{m}/\underline{z} 451 (M+H)^+]$  and a 2,4-decadienoic acid (<u>3</u>)  $[C_{10}H_{16}O_2; EI-MS, \underline{m}/\underline{z} 182 (M^+, methyl ester)].$ 

The <sup>1</sup>H and <sup>13</sup>C NMR spectra in DMSO-d<sub>6</sub> revealed that the compound <u>2</u> consists of a tetracene-quinone chromophore and an olivose moiety. Based on LSPD experiments, the structure of <u>2</u> was elucidated as shown in Fig. 3. Such a rearrangement as involved in the conversion of <u>1</u> with the modified benz[a]anthraquinone into the tetracene-quinone derivative (<u>2</u>) was reported in case of aquayamycin<sup>6</sup>, an isotetracenone antibiotic.





The compound <u>3</u> was determined to be  $(\underline{E},\underline{E})-2,4$ -decadienoic acid<sup>4)</sup> by the <sup>1</sup>H NMR spectral data in pyridine-d<sub>5</sub>. The geometrical configurations were elucidated by the coupling constants of olefinic protons  $(J_{2-H,3-H}=15.4Hz, J_{4-H}=15.0Hz)$ .

Treatment of <u>1</u> with 0.1 N HCl at 100°C for 10 min yielded a rearranged product (<u>4</u>)  $[C_{35}H_{36}O_9;$  FAB-MS, <u>m/z</u> 601 (M+H)<sup>+</sup>]. Comparison of the <sup>13</sup>C NMR spectrum<sup>5</sup>) of <u>4</u> with those of <u>2</u> and <u>3</u> indicates that the compound <u>4</u> consists of <u>2</u> and <u>3</u>. Since the acylation shift was observed on 4'-H ( $\delta$  2.90 + 4.67), the structure of <u>4</u> was determined to be a 4'-O-[(<u>E,E</u>)-2,4-decadienoy1] derivative of <u>2</u>.

Thus, it is concluded that the structure of capoamycin (1) is as shown in

Fig. 1 with the uncertainty about the stereochemistry of the ring juncture between C and D and the absolute configuration.

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## References and footnotes

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- 2) Y. Ichikawa, J. Cell. Physiol., <u>74</u>, 234 (1969).
- 3) <sup>1</sup>H and <sup>13</sup>C spectra were obtained on a JEOL GX-400 spectrometer operating at 400 MHz and 100 MHz, respectively. Chemical shifts are given in ppm using TMS as internal standard.
- 4)  ${}^{13}$ C NMR spectral data of <u>3</u> in CDCl<sub>3</sub> are as follows:  $\delta$  171.4 (C-1), 147.2 (C-3), 145.9 (C-5), 128.0 (C-4), 117.8 (C-2), 33.0 (C-6), 31.6 (C-8), 28.6 (C-7), 22.7 (C-9) and 14.3 (C-10).
- 5) <sup>13</sup>C NMR spectral data of  $\underline{4}$  in CDCl<sub>3</sub>-CD<sub>3</sub>OD are as follows:  $\delta$  187.5(C-7), 186.3 (C-12), 167.3 (C-1"), 162.3 (C-1), 158.3 (C-8), 155.3 (C-5), 146.0 (C-3"), 145.5 (C-5"), 141.3 (C-3), 136.3 (C-9), 132.7 (C-10), 132.5 (C-11a), 128.7 (C-1a), 127.7 (C-4"), 125.4 (C-6a), 124.7 (C-5a), 118.5 (C-11), 117.9 (C-2"), 117.5 (C-6), 116.4 (C-7a), 116.0 (C-4), 115.5 (C-2), 108.9 (C-12a), 78.3 (C-4'), 74.4 (C-5'), 71.3 (C-1' and C-3'), 39.9 (C-2'), 33.1 (C-6"), 31.4 (C-8"), 28.4 (C-7"), 22.6 (C-9"), 22.3 (3-CH<sub>3</sub>), 18.2 (C-6') and 14.0(C-10").
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